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Proficiency Test Report AQA 18-17 Cocaine

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I would like to thank the management and staff of the participating laboratories for supporting the study. It is only through widespread participation that we can provide an effective service to laboratories.

The assistance of the following NMI staff members in the planning, conduct and reporting of the study is acknowledged.

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SUMMARY

AQA 18-17 was conducted in December 2018. Three test samples of cocaine hydrochloride were sent to thirty-one laboratories. Three laboratories submitted extra sets of results analysed independently by different analysts.

The assigned values were the robust average of participants' results.

Traceability: The consensus of participants' results is not traceable to any external reference, so although expressed in SI units, metrological traceability has not been established.

The outcomes of the study were assessed against the aims as follows:

- *assess the proficiency of laboratories measuring cocaine in samples typical of a routine seizure;*

Laboratory performance was assessed by z-score and E_n -score.

Laboratories **1, 3, 5, 6, 7, 8, 9, 10, 11, 13, 14, 15, 16, 17, 18, 19, 20, 22, 23, 24, 26, 27, 28, 29, 31, 32, 33** and **34** returned satisfactory z and E_n -scores for all results.

Laboratory **25** returned questionable or unsatisfactory z-scores for all samples. Laboratory **2** returned unsatisfactory E_n -scores for all samples.

Of the 102 results for which z-scores were calculated, 91 (89%) returned $|z| \leq 2$ indicating a satisfactory performance.

Of the 102 results for which $|E_n|$ -scores were calculated, 95 (93%) returned $|E_n| \leq 1$ indicating agreement of the participants' results with the assigned value within their respective expanded uncertainties.

- *develop a practical application of traceability and measurement uncertainty and provide participants with information that will assist uncertainty estimates; and*

Ninety-nine results (97%) were reported with an associated expanded uncertainty. Laboratory **2** did not report uncertainty. This laboratory was not accredited.

Laboratories **12, 15, 20, 24, 30** and **33** reported an identical uncertainty for samples which were of significantly different concentrations.

The magnitude of reported uncertainties was within the range 1.5% to 52% relative.

- *test the ability of participants to identify a cutting agent commonly found in controlled drug preparation*

Samples were prepared using an illicit seizure of cocaine hydrochloride, approximately 84% base (m/m) supplied by the Australian Federal Police. The study coordinator added, phenacetin in Sample S2 and procaine in Sample S3.

Thirty-three participants (96%) reported on the identity of the cutting agents and correctly identified all of them.

1 INTRODUCTION

1.1 NMI Proficiency Testing Program

The National Measurement Institute (NMI) is responsible for Australia's national measurement infrastructure, providing a range of services including a chemical proficiency testing program.

Proficiency testing (PT) is: 'evaluation of participant performance against pre-established criteria by means of interlaboratory comparison.'¹ NMI PT studies target chemical testing in areas of high public significance such as trade, environment, law enforcement and food safety. NMI offers studies in:

- pesticide residues in fruit and vegetables, soil and water;
- petroleum hydrocarbons in soil and water;
- PFAS in water, soil and biota;
- metals in soil, water, food and pharmaceuticals;
- controlled drug assay and clandestine laboratory;
- allergens in food; and
- Folic acid in flour.

1.2 Study Aims

The aims of the study were to:

- assess the proficiency of laboratories measuring cocaine in samples typical of a routine seizure;
- develop a practical application of traceability and measurement uncertainty and provide participants with information that will assist uncertainty estimates; and
- Test the ability of participants to identify a cutting agent commonly found in controlled drug preparation.

The choice of the test method was left to the participating laboratories.

1.3 Study Conduct

NMI is accredited by the National Association of Testing Authorities, Australia (NATA) to ISO 17043¹ as a provider of proficiency testing schemes. This controlled drug proficiency test is within the scope of NMI's accreditation.

The conduct of NMI proficiency tests is described in the NMI Chemical Proficiency Testing Study Protocol.² The statistical methods used are described in the NMI Chemical Proficiency Testing Statistical Manual.³ These documents have been prepared with reference to ISO 17043 and The International Harmonized Protocol for Proficiency Testing of (Chemical) Analytical Laboratories.⁴

2 STUDY INFORMATION

2.1 Study Timetable

The timetable of the study was:

| | |
|------------------------|-------------------|
| Invitation issued: | 24 September 2018 |
| Samples dispatched: | 06 December 2018 |
| Results due: | 22 February 2019 |
| Interim report issued: | 01 March 2019 |

2.2 Participation

A total of ninety-nine international, national, state government and private laboratories were invited to participate.

Thirty-one laboratories agreed to participate and submitted results. Three laboratories requested two sets of test samples in order to be analysed by different analysts and reported two sets of results.

2.3 Test Material Specification

Three test samples were prepared in September 2018. The starting material was cocaine hydrochloride approximately 84% base (m/m) supplied by the Australian Federal Police. Phenacetin and procaine purchased from Sigma Aldrich were used as cutting agents. Sample S1 was uncut, phenacetin was used for Sample S2 and procaine for Sample S3.

The cocaine was ground and sieved through a 180 µm sieve. The cutting agents were processed similarly to the cocaine powder.

Test samples were then prepared by mixing a known mass of sieved drug material with a known mass of sieved cutting agent in a tumbler overnight.

Portions of 150 mg of each of the test samples were weighed into labelled glass vials.

Sample S1 was prepared to contain ~84% cocaine base (m/m).

Sample S2 was prepared to contain ~40% cocaine base (m/m).

Sample S3 was prepared to contain ~26% cocaine base (m/m).

2.4 Laboratory Code

Each participant was randomly assigned a confidential laboratory code.

2.5 Test Sample Homogeneity

The preparation of homogeneous test samples is an important part of a proficiency testing study. Given the small (<150 mg) test portions normally used for controlled substances analysis the particle size must be sufficiently small and uniformly distributed to ensure minimal influence on analytical precision.

The procedure for the preparation of the study samples has been validated in previous studies. No homogeneity testing was conducted in this proficiency study. Results returned by the participants gave no reason to question the homogeneity of test samples.

2.6 Sample Dispatch and Receipt

A set of three test samples, each containing approximately 150 mg of test material, were dispatched on 06 December 2019.

The following items were packaged with the samples:

- a covering letter with instructions for participants; and
- a form for participants to confirm the receipt of the test samples.

An Excel spreadsheet for the electronic reporting of results was e-mailed to participants.

2.7 Instructions to Participants

Participants were asked to analyse the samples using their routine quantitative method and return the following information:

- one result for each sample as % m/m cocaine base;
- an estimate of the expanded uncertainty associated with the result as % m/m cocaine base at the 95% confidence level;
- brief detail on how the uncertainty was calculated e.g. uncertainty budget method;
- the identity of the cutting agents in all three samples, if part of routine analysis;
- origin and stated purity of the analytical reference standard used;
- brief summary of the quantitative method used;
- the completed results sheet by 22 February 2019, as late results cannot be included in the report; and
- Any other comment.

2.8 Interim Report

An interim report was emailed to all participants on 01 March 2019.

3 PARTICIPANT LABORATORY INFORMATION

3.1 Test Method Summaries

Reported participant method summary is presented for information in Table 1.

Table 1 Participant Summary of Test Methods

| Lab. Code | Extraction solvent | Internal standard | Calib. points | Technique | Detector | Column |
|-----------|---|--|---------------|-------------------------|-----------------------|--|
| 1 | ACN/MEOH/H ₂ O | Analog of cocaine | 7 | UPLC | MSMS | C-18 coloumn |
| 2 | Ethanol | Propylparaben | 7 | UPLC | DAD | BEH Shield RP18 |
| 3 | Methanol | | 4 | HPLC | DAD | Zorbax Eclipse XDB-C18 |
| 4 | water/acetonitrile/n10 sulphuric acid 90:10:1 | | 3 | HPLC | Diode Array | Shimpack XR-ODS |
| 5 | | | | HPLC | DAD | ZORBAX ECLIPSE XDB-C18 (5mm pore size,4,6mmx150mm) |
| 6 | Methanol | Tetracosane | 4 | GC | FID | SGE 12 x 0,22 mm |
| 7 | Methanol | Vanillin | 1 | LC | DAD | Lichrospher 60 RP-select B, 25cm x 4 mm id, 5um |
| 8 | Acetonitrile/water 20:80 acidified | | 5 | HPLC | UV | C18 |
| 9 | Ethanol | 2,2,2-triphenylacetophenone (TPAP) | 3 | GC | FID, MSD | HP-1MS |
| 10 | Sodiumphosphate (pH4,5) | | 4 | HPLC | UV-DAD | Hypersil GOLD C8 |
| 11 | Mobile Phase (S1 and S3); Chloroform (S2) | None (S1 and S3); 2,2,2-triphenylacetophenone (S2) | 4 | HPLC (S1,S3) GC (S2) | PDA (S1, S3) FID (S2) | C18 ubondapack (S1 and S3); HP-5 (S2) |
| 12 | Acetonitrile:water 25:75 | | 3 | HPLC | UV | ODS2-interpak 25.0cm x 4.6mm |
| 13 | Dichloromethane | Tetracosane | 7 | GC | MS | DB5 |
| 14 | Acetonitril/water | | 1 | HPLC | DAD | Kromasil |
| 15 | Water/Acetonitrile (80:20) | | 3 | HPLC | UV/VIS | C18 |
| 16 | ethanol | tribenzylamine | 6 | GC | FID | HP5 |

| Lab. Code | Extraction solvent | Internal standard | Calib. points | Technique | Detector | Column |
|-----------|---|--------------------------------|---------------|-----------|-----------|---|
| 17 | CDCI3 | 1,4-bis(trimethylsilyl)benzene | | QNMR | | |
| 18 | Methanol | | 6 | UPLC | PDA | Acquity UPLC BEH 1.7um 2.1 x 100mm |
| 19 | Ethanol | Tetracosane | 6 | GC | FID | HP5 |
| 20 | Methanol | Diazepam | 6 | GC | FID | J&W 128-5512 |
| 21 | acétonitrile/water 80/20 | External Standard | 2 | HPLC | DAD | 09-D-29 |
| 22 | Acetonitrile | Strychnine | 6 | GC | FID | HP-1 |
| 23 | Ethanol | Tetracosane | 3 | GC | FID | BPX5 |
| 24 | 72% water ultra pure + 28% acétonitrile | | 5 | HPLC | UV | Kromasil C8 |
| 25 | Methanol | | 6 | UPLC | PDA | Acquity UPLC BEH 1.7um 2.1 x 100mm |
| 26 | Acetonitrile/Methanol (95:5) | Pholcodine 1mg/ml | 3 | UPLC | PDA | ACQUITY C-18 |
| 27 | Acetonitrile:Water 75:25 | Diethylphthalate | 3 | UPLC | DAD | BEH C18 1.7mm (2.1x100mm) |
| 28 | ethanol | tribenzylamine | 4- | GC | FID | HP-1 |
| 29 | acétonitrile/water (80/20) | | 3 | HPLC | DAD | C8 |
| 30 | Ethanol | Eicosane | 1 | GC | MS | ZB -5ms |
| 31 | Methanol | | 5 | HPLC | DAD | Kinetex 2.6 µ XB-C18 |
| 32 | Methanol | External Standard | 6 | HPLC | UV 235 nm | Phenomenex C18 5um Luna |
| 33 | Acetonitrile:Water (40:60) | | 5 | HPLC | UV | KROMASIL |
| 34 | Methanol | Vanillin | 1 | LC | DAD | Lichrospher 60 RP-select B, 25cm x 4 mm id, 5um |

3.2 Reported Basis of Participants' Measurement Uncertainty Estimates

Participant returns as received are listed in Table 2.

Table 2 Reported Basis of Uncertainty Estimate

| Lab. Code | Approach to Estimating MU | Information Sources for MU Estimation | | Guide Document for Estimating MU |
|-----------|--|---------------------------------------|---|-----------------------------------|
| | | Precision | Method Bias | |
| 1 | Top Down - precision and estimates of the method and laboratory bias | Control samples | | |
| 2 | | | | |
| 3 | | Control samples, Duplicate analysis | Standard purity | Eurolab Technical Report No1/2007 |
| 4 | Professional judgment | Control samples, Duplicate analysis | Standard purity, Instrument calibration | ISO/GUM |
| 5 | Top Down - precision and estimates of the method and laboratory bias | Control samples, Duplicate analysis | Laboratory bias from PT studies, Standard purity Instrument calibration, Masses and volumes, Homogeneity of sample | Nordtest Report TR537 |
| 6 | Top Down - precision and estimates of the method and laboratory bias | Control samples | Laboratory bias from PT studies, Recoveries of spiked samples, Standard purity, Matrix effects, Instrument calibration, Masses and volumes, Homogeneity of sample | EA-4/16: 2003 and ILAG G-17:2002 |
| 7 | Top Down - precision and estimates of the method and laboratory bias | Control samples, Duplicate analysis | Laboratory bias from PT studies, Recoveries of spiked samples | Eurachem/CITAC Guide |
| 8 | Top Down - precision and estimates of the method and laboratory bias | Control samples, Duplicate analysis | Recoveries of spiked samples | Eurachem/CITAC Guide |
| 9 | Bottom Up (ISO/GUM, fish bone/ cause and effect diagram) | Control samples, Duplicate analysis | Recoveries of spiked samples, Standard purity, Matrix effects, Instrument calibration, Masses and volumes, Homogeneity of sample | Eurachem/CITAC Guide |
| 10 | Top Down - precision and estimates of the method and laboratory bias | Control samples, Duplicate analysis | Laboratory bias from PT studies | Nordtest Report TR537 |
| 11 | | | | |
| 12 | Professional judgment | | Instrument calibration | ISO/GUM |

| Lab. Code | Approach to Estimating MU | Information Sources for MU Estimation | | Guide Document for Estimating MU |
|-----------|---|---|---|----------------------------------|
| | | Precision | Method Bias | |
| 13 | Top Down - precision and estimates of the method and laboratory bias | Control samples, Duplicate analysis | Laboratory bias from PT studies, Recoveries of spiked samples, Standard purity, Instrument calibration | |
| 14 | Standard deviation of replicate analyses multiplied by 2 or 3 | Control samples | | |
| 15 | Standard deviation of replicate analyses multiplied by 2 or 3 | Control samples, Duplicate analysis | Standard purity, Masses and volumes, Homogeneity of sample | Eurachem/CITAC Guide |
| 16 | Standard deviation of replicate analyses multiplied by 2 or 3 | Control samples | Standard purity | |
| 17 | Top Down - precision and estimates of the method and laboratory bias | Control samples, Duplicate analysis | Matrix effects, Instrument calibration | Eurachem/CITAC Guide |
| 18 | Top Down - precision and estimates of the method and laboratory bias | Control samples, Duplicate analysis | Standard purity, Instrument calibration, Masses and volumes, Homogeneity of sample | Nata Technical Note 33 |
| 19 | Top Down - precision and estimates of the method and laboratory bias | Control samples, Duplicate analysis | Standard purity, Matrix effects | ISO/GUM |
| 20 | Estimating Measurement Uncertainty by black box by pairs of values | Standard deviation from PT studies only | | ISO/GUM |
| 21 | | Duplicate analysis | Standard purity, Instrument calibration | ISO/GUM |
| 22 | Top Down - reproducibility (standard deviation) from PT studies used directly | Control samples, Duplicate analysis | Laboratory bias from PT studies, Standard purity, Instrument calibration, Masses and volumes, Homogeneity of sample | Nata Technical Note 33 |
| 23 | Uncertainty budget | Control samples, Duplicate analysis | Standard purity, Instrument calibration, Masses and volumes | Internal SOP |
| 24 | Bottom Up (ISO/GUM, fish bone/ cause and effect diagram) | Control samples, Duplicate analysis | Laboratory bias from PT studies, Standard purity, Instrument calibration, Masses and volumes | ISO/GUM |
| 25 | Top Down - precision and estimates of the method and laboratory bias | Control samples, Duplicate analysis | Standard purity, Instrument calibration, Masses and volumes, Homogeneity of sample | Nata Technical Note 33 |
| 26 | Top Down - precision and estimates of the method and laboratory bias | Control samples, Duplicate analysis | Standard purity, Instrument calibration, Masses and volumes, Homogeneity of sample | Nata Technical Note 33 |

| Lab. Code | Approach to Estimating MU | Information Sources for MU Estimation | | Guide Document for Estimating MU |
|-----------|--|---------------------------------------|---|---|
| | | Precision | Method Bias | |
| 27 | Top Down - precision and estimates of the method and laboratory bias | Control samples, Duplicate analysis | Standard purity, Homogeneity of sample | Eurachem/CITAC Guide |
| 28 | Top Down - precision and estimates of the method and laboratory bias | Control samples, Duplicate analysis | Laboratory bias from PT studies | Internal quality online document based on Eurachem/CITAC, ISO/GUM |
| 29 | Top Down - precision and estimates of the method and laboratory bias | Control samples | Laboratory bias from PT studies , Standard purity | |
| 30 | Standard deviation of replicate analyses multiplied by 2 or 3 | Control samples, Duplicate analysis | Masses and volumes, Homogeneity of sample | Eurachem/CITAC Guide |
| 31 | Bottom Up (ISO/GUM, fish bone/ cause and effect diagram) | Control samples | Standard purity, Instrument calibration | Eurachem/CITAC Guide |
| 32 | Top Down - precision and estimates of the method and laboratory bias | Control samples | Laboratory bias from PT studies | Eurachem/CITAC Guide |
| 33 | Top Down - precision and estimates of the method and laboratory bias | Control samples, Duplicate analysis | Instrument calibration, Masses and volumes | |
| 34 | Top Down - precision and estimates of the method and laboratory bias | Control samples, Duplicate analysis | Laboratory bias from PT studies, Recoveries of spiked samples | Eurachem/CITAC Guide |

3.3 Details of Participant Calibration Standard

Participant returns as received are listed in Table 3.

Table 3 Participant Calibration Standard

| Lab. Code | Reference Standard* | Purity (%) |
|-----------|-------------------------|----------------|
| 1 | Unikem | 100 |
| 2 | NMI | 96.1 |
| 3 | LGC | 1 ±0,003 mg/mL |
| 4 | LGC | 99.7 |
| 5 | LIPOMED | 99.35 |
| 6 | Merck | 100 |
| 7 | Lipomed | 99.706 ± 0.007 |
| 8 | Johnson Matthey | 99.7 |
| 9 | NMI | 96.1 +/- 2.6 |
| 10 | Sigma Aldrich | 100 |
| 11 | Macfarlan Smith Limited | 100.4 |
| 12 | MacFarlane Smith | 99.1 |
| 13 | Lipomed | 86.7 |
| 14 | Sigma-Aldrich | 98.7 |
| 15 | Sigma-Aldrich | 99.2 |
| 16 | NMI | 99.8 |
| 17 | | |
| 18 | NMI | 96.1 |
| 19 | Alcaliber | 100.1 |
| 20 | LIPOMED | 99.7 |
| 21 | NMI | 96.1 |
| 22 | NMI | 96.1 |
| 23 | NMI | 96.1 |
| 24 | | |
| 25 | NMI | 96.1 |
| 26 | NMI | 96.1 |
| 27 | NMI | 99.8 ± 2.0 |
| 28 | Fagron | 99.5 |
| 29 | Lipomed | 99.706 |
| 30 | Sigma-aldrich | 99 |
| 31 | Lipomed | >98.5 |
| 32 | | 100 |
| 33 | SIGMA | |
| 34 | Lipomed | 99.706 ± 0.007 |

3.4 Participants' Comments

The study manager welcomes comments or suggestions from participants as it provides information which will improve future studies. All returns are listed as received in Table 4 along with the study manager's response, where appropriate.

Table 4 Participant Comments

| Lab. Code | Participant comments | Study Manager's response |
|-----------|---|--|
| 5 | Qualitative analysis was carried out by GC-MS | |
| 11 | Insufficient sample to repeat analysis if needed. | Most participants use less than 50 mg for each analysis. For reasons of security and accountability, NMI conducts these PT's using the minimum practical amount of drug. |
| 24 | Solutions 1 and 2 were analyzed in HPLC-UV. Solution 3 was analyzed in GCMS | |

4 PRESENTATION OF RESULTS AND STATISTICAL ANALYSIS

4.1 Results Summary

Participant results are listed in Tables 5 to 7 with resultant summary statistics: mean, median, maximum, minimum, robust average, robust standard deviation (Robust SD) and robust coefficient of variation (Robust CV).

Bar charts of results and performance scores are presented in Figures 2 to 4.

An example chart with interpretation guide is shown in Figure 1.

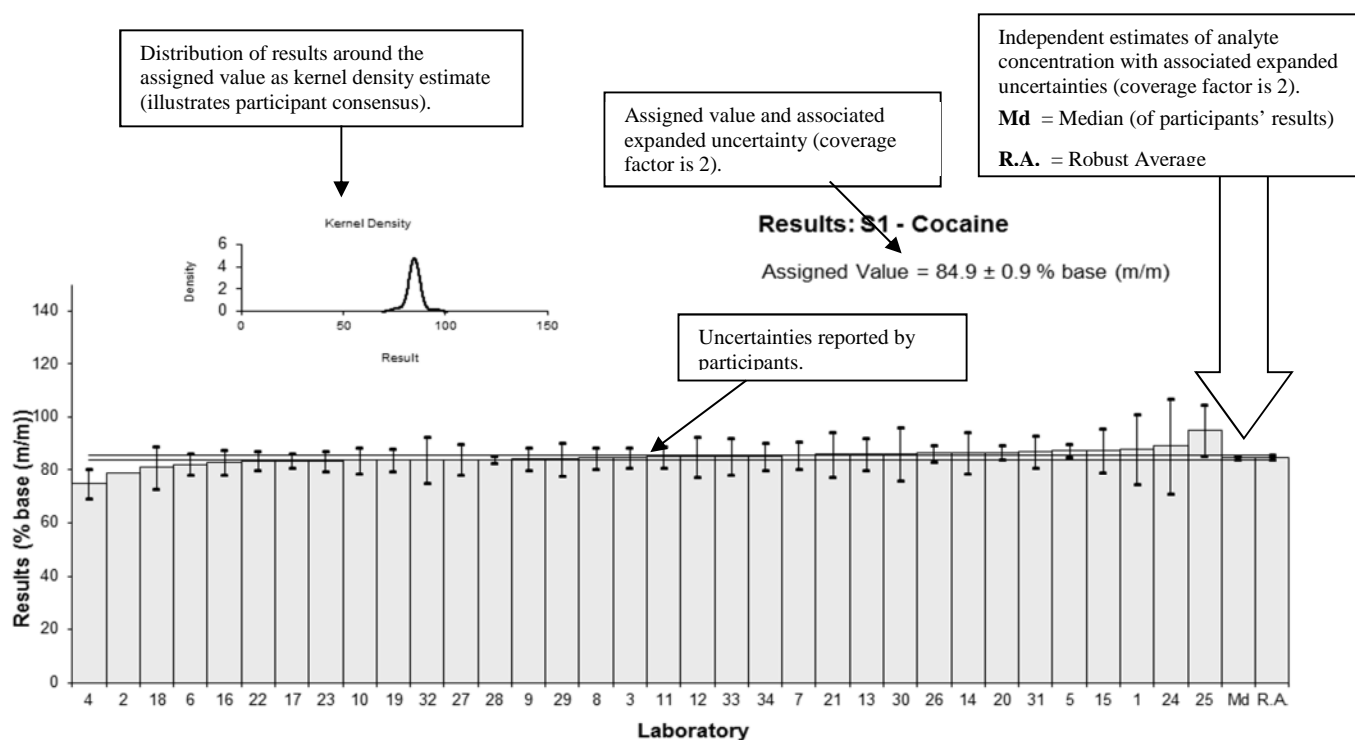


Figure 1 Guide to Presentation of Results

4.2 Assigned Value

The assigned value is defined as: 'value attributed to a particular property of a proficiency test item.'¹

For a proficiency test, the assigned value is the best available measurement of the true concentration of an analyte in the test sample.

4.3 Performance Coefficient of Variation (PCV)

The performance coefficient of variation (PCV) is a measure of the between laboratory variation that in the judgement of the study organiser would be expected from participants given the sample concentration. It is important to note that this is a performance measure set by the study coordinator; it is not the coefficient of variation of participant results.

4.4 Target Standard Deviation

The target standard deviation (σ) is the product of the assigned value (X) and the performance laboratory coefficient of variation (PCV) as presented in Equation 1. This value is used for calculation of participant z-score.

$$\sigma = X * PCV \quad \text{Equation 1}$$

4.5 z-Score

For each participant result a z-score is calculated according to Equation 2 below:

$$z = \frac{(\chi - X)}{\sigma} \quad \text{Equation 2}$$

where:

- z is z-score
- χ is participants' result
- X is the study assigned value
- σ is the target standard deviation from equation 1

A z-score with absolute value ($|z|$):

- $|z| \leq 2$ is satisfactory;
- $2 < |z| < 3$ is questionable;
- $|z| \geq 3$ is unsatisfactory.

4.6 E_n-Score

The E_n-score is complementary to the z-score in assessment of laboratory performance.

E_n-score includes measurement uncertainty and is calculated according to Equation 3 below:

$$E_n = \frac{(\chi - X)}{\sqrt{U_\chi^2 + U_X^2}} \quad \text{Equation 3}$$

where:

- E_n is E_n-score
- χ is a participants' result
- X is the assigned value
- U_χ is the expanded uncertainty of the participant's result
- U_X is the expanded uncertainty of the assigned value

An E_n-score with absolute value ($|E_n|$):

- $|E_n| \leq 1$ is satisfactory;
- $|E_n| > 1$ is unsatisfactory.

4.7 Traceability and Measurement Uncertainty

Laboratories accredited to ISO/IEC Standard 17025:2017⁵ must establish and demonstrate the traceability and measurement uncertainty associated with their test results. Guidelines for quantifying uncertainty in analytical measurement are described in the Eurachem /CITAC Guide.⁶

5 TABLES AND FIGURES

Table 5

Sample Details

| | |
|-------------------|--------------|
| Sample No. | S1 |
| Matrix. | Powder |
| Analyte. | Cocaine |
| Units | % base (m/m) |

Participant Results

| Lab Code | Result | Uncertainty | z-Score | E _n -Score |
|----------|--------|-------------|---------|-----------------------|
| 1 | 88 | 13.2 | 1.22 | 0.23 |
| 2 | 78.9 | NR | -2.36 | -6.67 |
| 3 | 84.7 | 4.0 | -0.08 | -0.05 |
| 4 | 75 | 5.6 | -3.89 | -1.75 |
| 5 | 87.3 | 2.4 | 0.94 | 0.94 |
| 6 | 82.2 | 4.2 | -1.06 | -0.63 |
| 7 | 85.5 | 5.2 | 0.24 | 0.11 |
| 8 | 84.5 | 4.2 | -0.16 | -0.09 |
| 9 | 84.1 | 4.3 | -0.31 | -0.18 |
| 10 | 83.6 | 5.0 | -0.51 | -0.26 |
| 11 | 85 | 4.1 | 0.04 | 0.02 |
| 12 | 85 | 7.5 | 0.04 | 0.01 |
| 13 | 86 | 6 | 0.43 | 0.18 |
| 14 | 86.5 | 7.8 | 0.63 | 0.20 |
| 15 | 87.4 | 8.166 | 0.98 | 0.30 |
| 16 | 82.9 | 4.7 | -0.79 | -0.42 |
| 17 | 83.5 | 2.8 | -0.55 | -0.48 |
| 18 | 81 | 8.1 | -1.53 | -0.48 |
| 19 | 83.8 | 4.4 | -0.43 | -0.24 |
| 20 | 86.6 | 2.6 | 0.67 | 0.62 |
| 21 | 85.9 | 8.6 | 0.39 | 0.12 |
| 22 | 83.4 | 3.6 | -0.59 | -0.40 |
| 23 | 83.5 | 3.8 | -0.55 | -0.36 |
| 24 | 89 | 18 | 1.61 | 0.23 |
| 25 | 95 | 9.5 | 3.97 | 1.06 |
| 26 | 86.3 | 3 | 0.55 | 0.45 |
| 27 | 84.0 | 6.0 | -0.35 | -0.15 |
| 28 | 84.0 | 1.3 | -0.35 | -0.57 |
| 29 | 84.20 | 6.32 | -0.27 | -0.11 |
| 30 | 86.08 | 10 | 0.46 | 0.12 |
| 31 | 87 | 6.1 | 0.82 | 0.34 |
| 32 | 83.8 | 8.6 | -0.43 | -0.13 |
| 33 | 85 | 7 | 0.04 | 0.01 |
| 34 | 85.0 | 5.1 | 0.04 | 0.02 |

Statistics

| | | |
|-----------------------|------|-----|
| Assigned Value | 84.9 | 0.9 |
| Robust Average | 84.9 | 0.9 |
| Median | 84.9 | 0.7 |
| Mean | 84.8 | |
| N | 34 | |
| Max. | 95 | |
| Min. | 75 | |
| Robust SD | 2.0 | |
| Robust CV | 2.4% | |

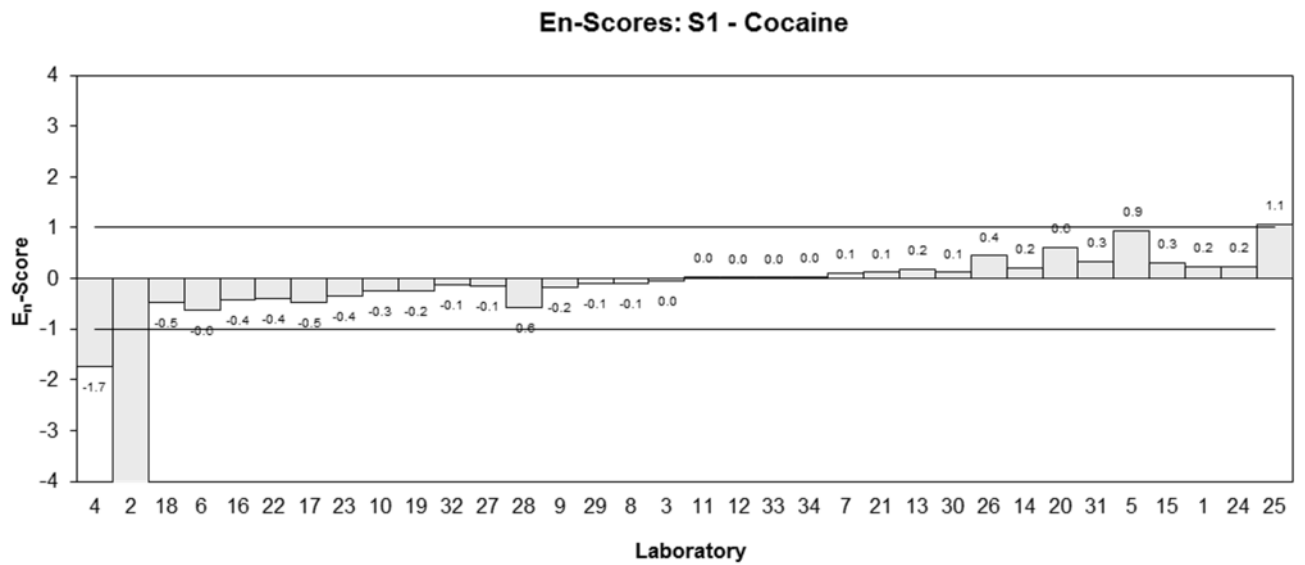
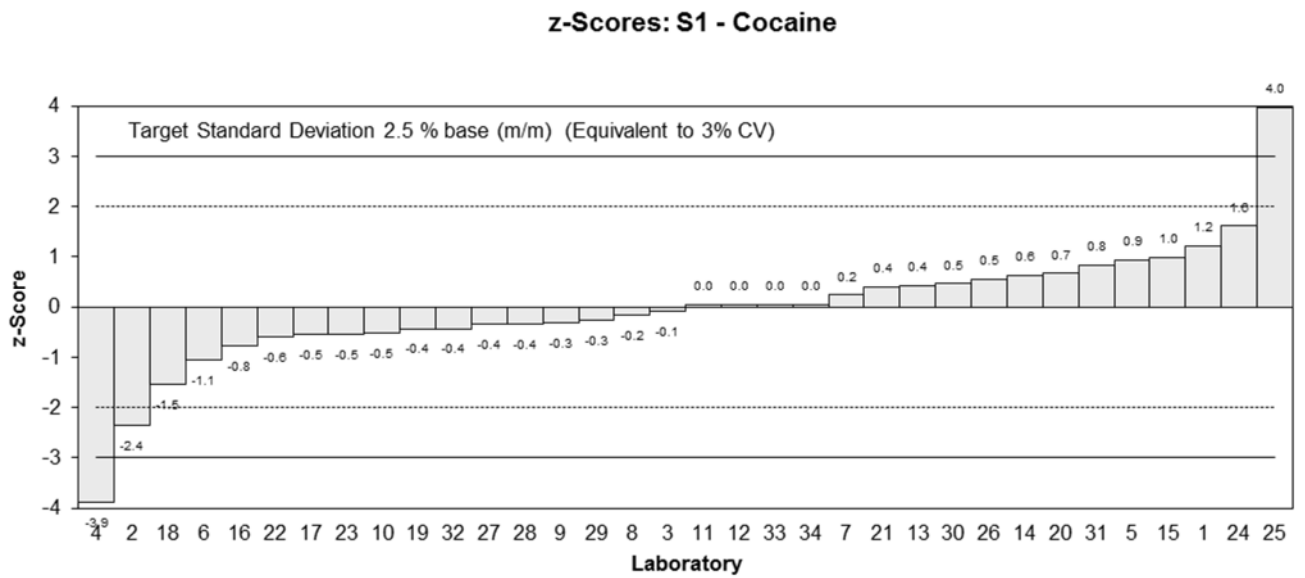
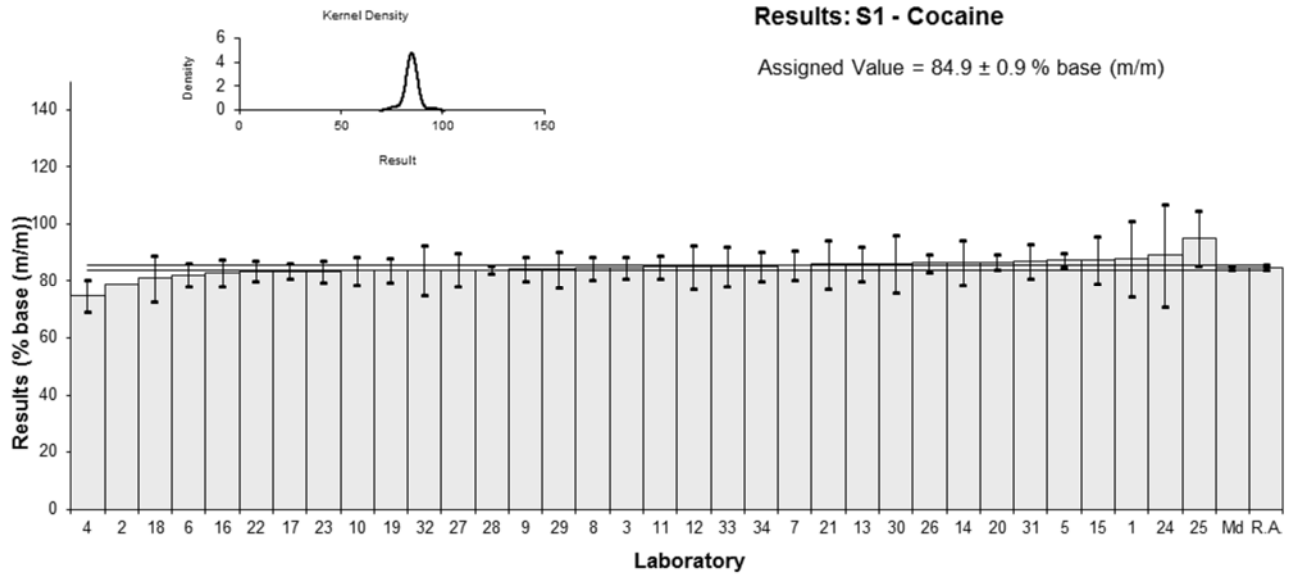


Figure 2

Table 6

Sample Details

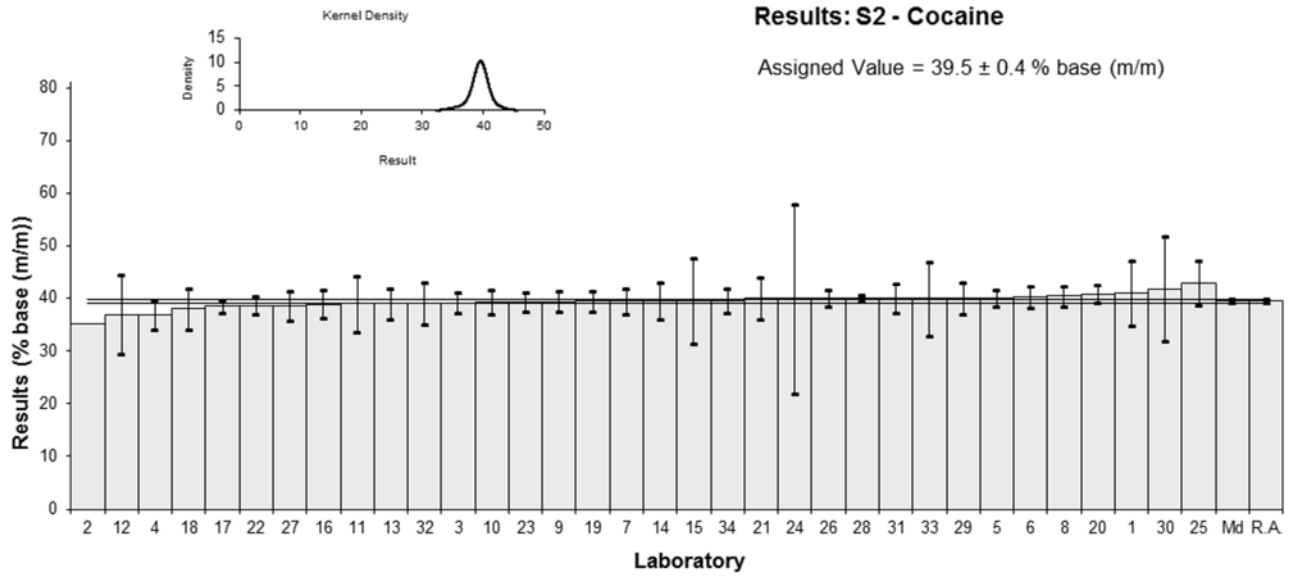
| | |
|-------------------|--------------|
| Sample No. | S2 |
| Matrix. | Powder |
| Analyte. | Cocaine |
| Units | % base (m/m) |

Participant Results

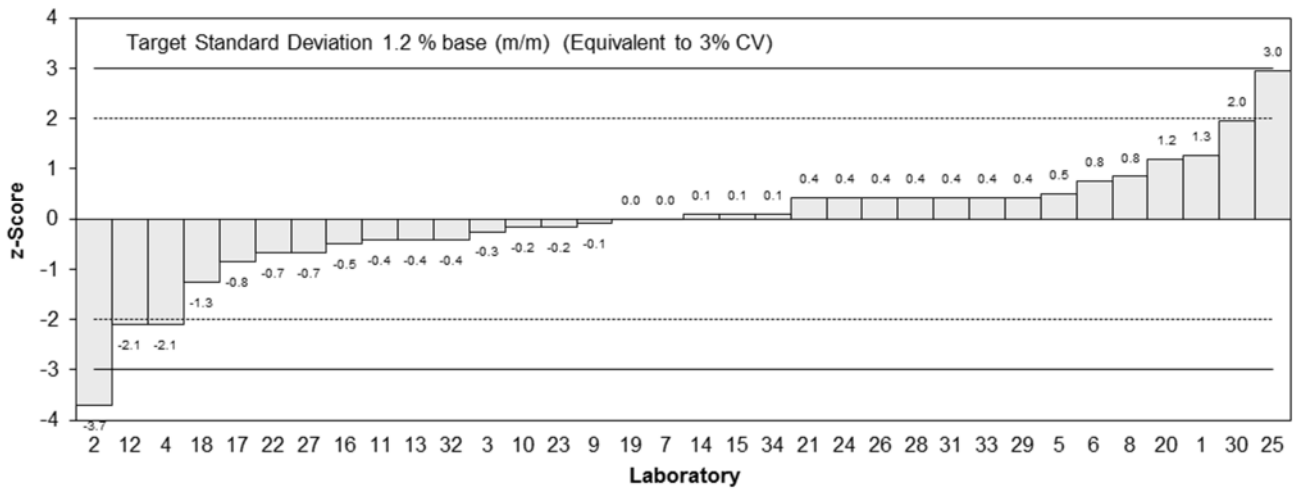
| Lab Code | Result | Uncertainty | z-Score | E _n -Score |
|----------|--------|-------------|---------|-----------------------|
| 1 | 41 | 6.2 | 1.27 | 0.24 |
| 2 | 35.1 | NR | -3.71 | -11.00 |
| 3 | 39.2 | 2.0 | -0.25 | -0.15 |
| 4 | 37 | 2.8 | -2.11 | -0.88 |
| 5 | 40.1 | 1.5 | 0.51 | 0.39 |
| 6 | 40.4 | 2.1 | 0.76 | 0.42 |
| 7 | 39.5 | 2.4 | 0.00 | 0.00 |
| 8 | 40.5 | 2.0 | 0.84 | 0.49 |
| 9 | 39.4 | 2.0 | -0.08 | -0.05 |
| 10 | 39.3 | 2.3 | -0.17 | -0.09 |
| 11 | 39 | 5.4 | -0.42 | -0.09 |
| 12 | 37 | 7.5 | -2.11 | -0.33 |
| 13 | 39 | 3 | -0.42 | -0.17 |
| 14 | 39.6 | 3.6 | 0.08 | 0.03 |
| 15 | 39.6 | 8.166 | 0.08 | 0.01 |
| 16 | 38.9 | 2.7 | -0.51 | -0.22 |
| 17 | 38.5 | 1.3 | -0.84 | -0.74 |
| 18 | 38 | 3.8 | -1.27 | -0.39 |
| 19 | 39.5 | 2.0 | 0.00 | 0.00 |
| 20 | 40.9 | 1.7 | 1.18 | 0.80 |
| 21 | 40.0 | 4.0 | 0.42 | 0.12 |
| 22 | 38.7 | 1.7 | -0.68 | -0.46 |
| 23 | 39.3 | 1.8 | -0.17 | -0.11 |
| 24 | 40 | 18 | 0.42 | 0.03 |
| 25 | 43 | 4.3 | 2.95 | 0.81 |
| 26 | 40.0 | 1.6 | 0.42 | 0.30 |
| 27 | 38.7 | 2.8 | -0.68 | -0.28 |
| 28 | 40.0 | 0.6 | 0.42 | 0.69 |
| 29 | 40.01 | 3.00 | 0.43 | 0.17 |
| 30 | 41.82 | 10 | 1.96 | 0.23 |
| 31 | 40 | 2.8 | 0.42 | 0.18 |
| 32 | 39.0 | 4.0 | -0.42 | -0.12 |
| 33 | 40 | 7 | 0.42 | 0.07 |
| 34 | 39.6 | 2.4 | 0.08 | 0.04 |

Statistics

| | | |
|-----------------------|------|-----|
| Assigned Value | 39.5 | 0.4 |
| Robust Average | 39.5 | 0.4 |
| Median | 39.6 | 0.3 |
| Mean | 39.5 | |
| N | 34 | |
| Max. | 43 | |
| Min. | 35.1 | |
| Robust SD | 1.0 | |
| Robust CV | 2.5% | |



z-Scores: S2 - Cocaine



En-Scores: S2 - Cocaine

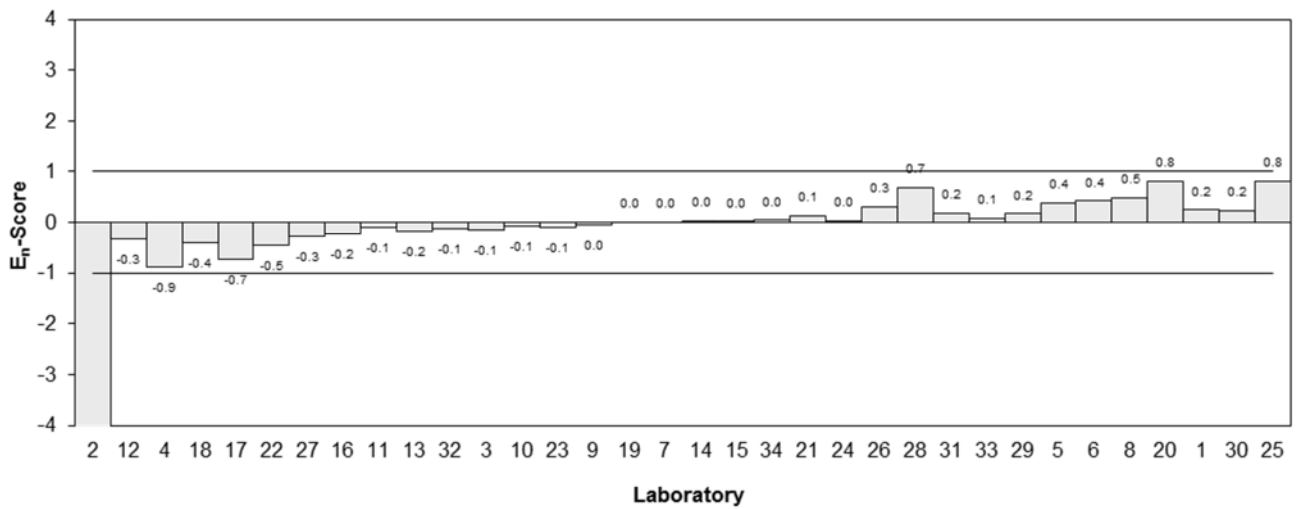


Figure 3

Table 7

Sample Details

| | |
|-------------------|--------------|
| Sample No. | S3 |
| Matrix. | Powder |
| Analyte. | Cocaine |
| Units | % base (m/m) |

Participant Results

| Lab Code | Result | Uncertainty | z-Score | E _n -Score |
|----------|--------|-------------|---------|-----------------------|
| 1 | 28 | 4.2 | 1.49 | 0.28 |
| 2 | 25.5 | NR | -1.62 | -3.25 |
| 3 | 27.0 | 1.0 | 0.25 | 0.19 |
| 4 | 28 | 2.1 | 1.49 | 0.56 |
| 5 | 26.3 | 2.2 | -0.62 | -0.22 |
| 6 | 27.1 | 1.4 | 0.37 | 0.21 |
| 7 | 26.7 | 1.7 | -0.12 | -0.06 |
| 8 | 26.4 | 2.6 | -0.50 | -0.15 |
| 9 | 26.3 | 1.4 | -0.62 | -0.34 |
| 10 | 26.2 | 1.6 | -0.75 | -0.36 |
| 11 | 26 | 1.2 | -1.00 | -0.63 |
| 12 | 25 | 7.5 | -2.24 | -0.24 |
| 13 | 27 | 2 | 0.25 | 0.10 |
| 14 | 27.3 | 2.5 | 0.62 | 0.20 |
| 15 | 26.9 | 8.166 | 0.12 | 0.01 |
| 16 | 25.9 | 1.8 | -1.12 | -0.49 |
| 17 | 25.9 | 0.9 | -1.12 | -0.91 |
| 18 | 27 | 2.7 | 0.25 | 0.07 |
| 19 | 28.4 | 1.5 | 1.99 | 1.03 |
| 20 | 26.9 | 1.7 | 0.12 | 0.06 |
| 21 | 24.6 | 2.5 | -2.74 | -0.87 |
| 22 | 26.0 | 1.1 | -1.00 | -0.68 |
| 23 | 26.9 | 1.3 | 0.12 | 0.07 |
| 24 | 27 | 14 | 0.25 | 0.01 |
| 25 | 32 | 3.2 | 6.47 | 1.61 |
| 26 | 27.4 | 1.5 | 0.75 | 0.39 |
| 27 | 26.5 | 2.0 | -0.37 | -0.15 |
| 28 | 26.8 | 0.4 | 0.00 | 0.00 |
| 29 | 26.86 | 2.01 | 0.07 | 0.03 |
| 30 | 28.88 | 10 | 2.59 | 0.21 |
| 31 | 28 | 1.7 | 1.49 | 0.69 |
| 32 | 26.9 | 2.8 | 0.12 | 0.04 |
| 33 | 28 | 6 | 1.49 | 0.20 |
| 34 | 26.2 | 1.6 | -0.75 | -0.36 |

Statistics

| | | |
|-----------------------|------|-----|
| Assigned Value | 26.8 | 0.4 |
| Robust Average | 26.8 | 0.4 |
| Median | 26.9 | 0.3 |
| Mean | 26.9 | |
| N | 34 | |
| Max. | 32 | |
| Min. | 24.6 | |
| Robust SD | 0.90 | |
| Robust CV | 3.4% | |

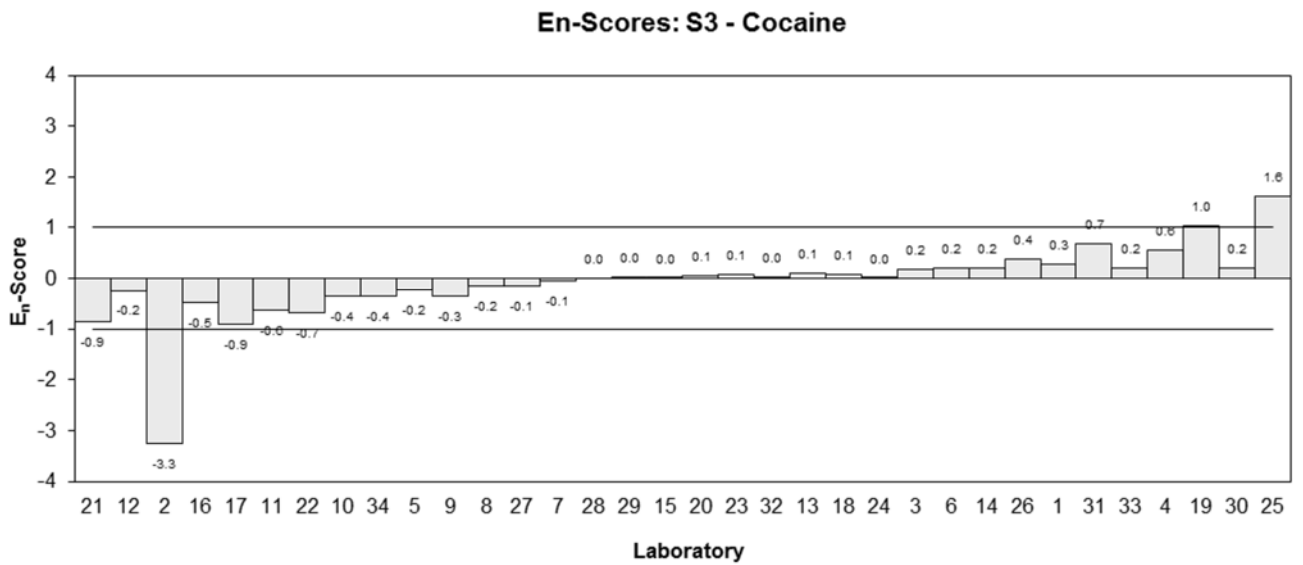
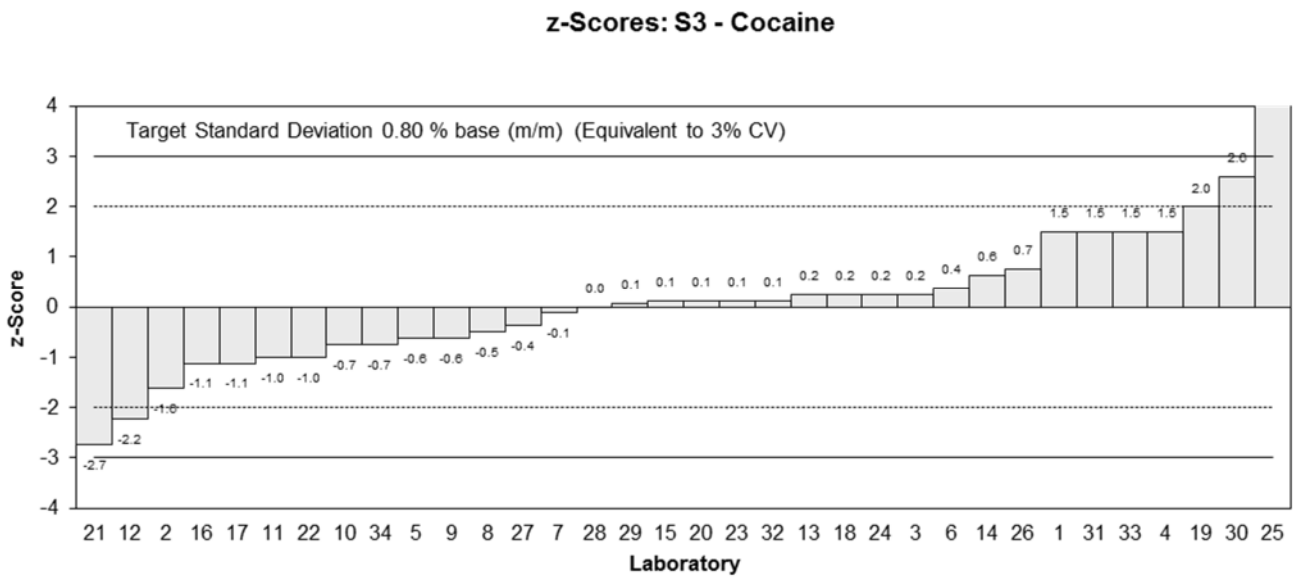
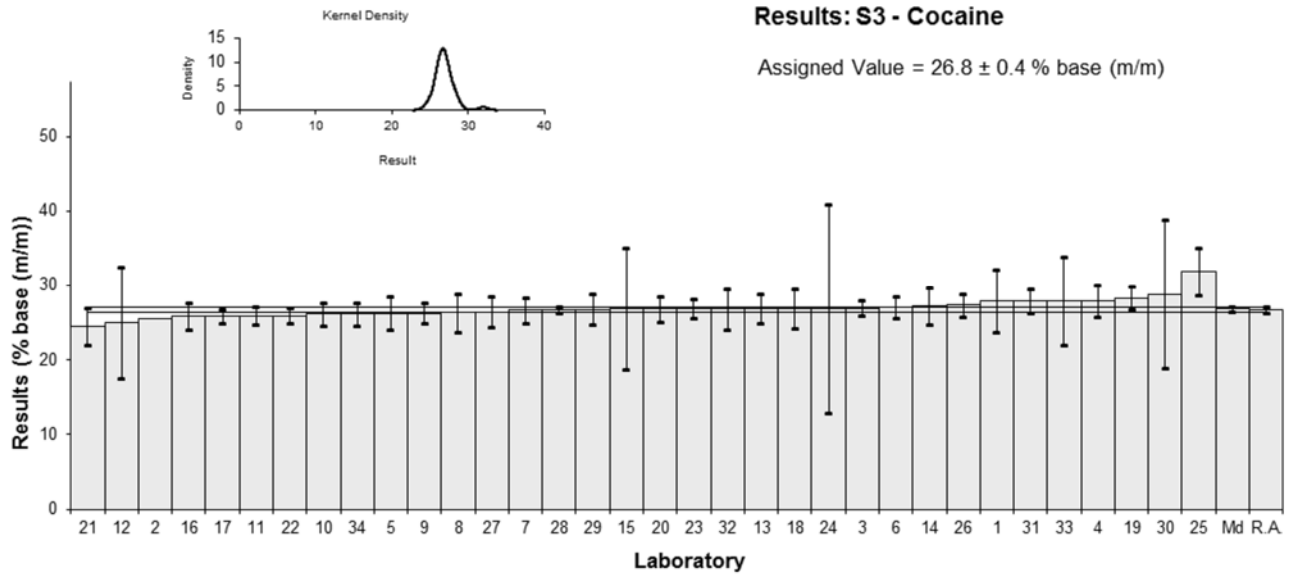


Figure 4

Table 8 Participants' identification of cutting agents

| Lab Code | Cutting agents | |
|----------|--------------------|------------------------|
| | S2 | S3 |
| 1 | Phenacetin | Procaine |
| 2 | Phenacetin :56,1 % | Procaine : 52,7 % |
| 3 | Phenacetin | Procaine |
| 4 | Phenacetin | Procaine |
| 5 | Phenacetine | Procaine |
| 6 | Phenacetine | Procaine |
| 7 | Phenacetin | Procaine |
| 8 | Phenacetin | Procaine |
| 9 | Phenacetin | Procaine |
| 10 | Phenacetin | Procaine |
| 11 | Phenacetin | procaine |
| 12 | Phenacetin | Procaine |
| 13 | Phenacetin | Procaine |
| 14 | Phenacetin | Procaine |
| 15 | Phenacetin | Procaine Hydrochloride |
| 16 | 52.7% phenacetin | Procaine |
| 17 | Phenacetin | Procaine |
| 18 | Phenacetin | Procaine |
| 19 | Phenacetin | Procaine |
| 20 | Phenacetin | Procaine |
| 21 | Phenacetin | Procaine |
| 22 | Phenacetin | Procaine |
| 23 | Phenacetin | Procaine |
| 24 | Phenacetin | Procaine |
| 25 | Phenacetin | Procaine |
| 26 | Phenacetin | procaine |
| 27 | Phenacetin | Procaine |
| 28 | Phenacetin | Procaine |
| 29 | Phenacetin | Procaine |
| 30 | Phenacetin | Procaine Hydrochloride |
| 31 | Phenacetin | Procaine |
| 32 | - | - |
| 33 | Phenacetin | - |
| 34 | Phenacetin | Procaine |

6 DISCUSSION OF RESULTS

6.1 Assigned Value

The assigned value is the robust average of the results reported by the participants. The robust average and associated expanded uncertainties were calculated using the procedure described in 'ISO13528:2015, Statistical methods for use in proficiency testing by interlaboratory comparisons'.⁷ The calculation procedure for the expanded uncertainty in Sample S1 is presented in Appendix 1.

Traceability: The consensus of participants' results is not traceable to any external reference, so although expressed in SI units, metrological traceability has not been established.

6.2 Measurement Uncertainty Reported by Participants

Participants were asked to report an estimate of the expanded measurement uncertainty associated with their results and the basis of this uncertainty estimate (Table 2).

It is a requirement of the ISO Standard 17025⁵ that laboratories have procedures to estimate the uncertainty of chemical measurements and to report this uncertainty in specific circumstances, including 'when the client's instruction so requires.' From 1 July 2012 this is also a requirement of ASCLD/Lab-International accreditation program.

Ninety-nine results (97%) were reported with an associated expanded uncertainty. Laboratory 2 did not report uncertainty. This laboratory was not accredited.

Laboratories 12, 15, 20, 24, 30 and 33 reported an identical uncertainty for samples which were of significantly different concentrations.

The magnitude of reported uncertainties was within the range 1.5% to 52% relative.

Sixty-seven of ninety-nine (68%) expanded uncertainties were between 3% and 10% relative to the result. Laboratories reporting uncertainties smaller than 3% or larger than 10% relative may wish to consider whether these estimates are realistic or fit for purpose.

Laboratories having a satisfactory z-score and an unsatisfactory E_n -score are likely to have underestimated the expanded uncertainty associated with the result.

In some cases the results were reported with an inappropriate number of significant figures. The recommended format is to write the uncertainty to no more than two significant figures and then to write the result with the corresponding number of decimal places (for example instead of $87.4 \pm 8.166\%$ the recommended format is $87.4 \pm 8.2\%$).⁶

6.3 z-Score

A target standard deviation equivalent to 3% performance coefficient of variation (PCV) was used to calculate z-scores. Target SDs, the between-laboratory coefficient of variation predicted by Thomson - Horwitz equation⁸ and between-laboratories coefficient of variation obtained in this study are presented in Table 9.

Table 9 Target standard deviations, coefficient of variations from predictive model and between laboratories

| Sample | Analyte | Assigned value (% base m/m) | Target SD (as PCV) | Thompson Horwitz CV | Between laboratories CV |
|--------|---------|-----------------------------|--------------------|---------------------|-------------------------|
| S1 | Cocaine | 84.9 | 3% | 2.0% | 2.4% |
| S2 | Cocaine | 39.5 | 3% | 2.3% | 2.5% |
| S3 | Cocaine | 26.8 | 3% | 2.4% | 3.4% |

A summary of z-scores by laboratory is presented in Figure 5.

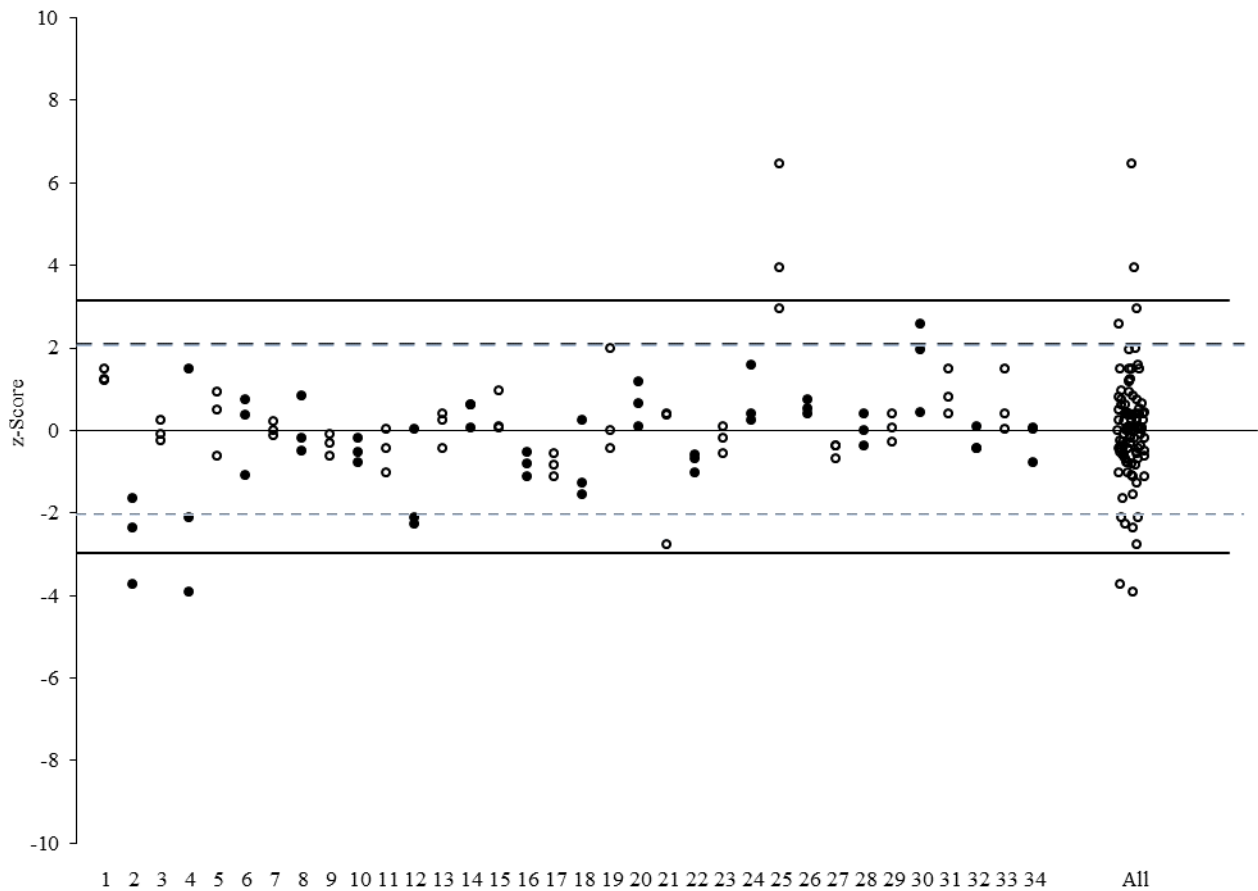


Figure 5 Summary of participants' z-score.

Ninety-one of 102 numeric results (89%) returned a satisfactory z-score with $|z| \leq 2$.

- Twenty-eight participants (82%) - **1, 3, 5, 6, 7, 8, 9, 10, 11, 13, 14, 15, 16, 17, 18, 19, 20, 22, 23, 24, 26, 27, 28, 29, 31, 32, 33** and **34** returned satisfactory scores for all three samples;
- Five participants returned at least one questionable or unsatisfactory z-score;
- Laboratory **25** returned questionable or unsatisfactory z-scores for all test samples demonstrating an unsatisfactory performance. This laboratory reported all results higher than the assigned value (positive bias) and may need to investigate the source of bias.

6.4 E_n-Score

The dispersal of participants' E_n-scores is graphically presented in Figure 6. Where a laboratory did not report an expanded uncertainty with a result, an expanded uncertainty of zero (0) was used to calculate the E_n-score.

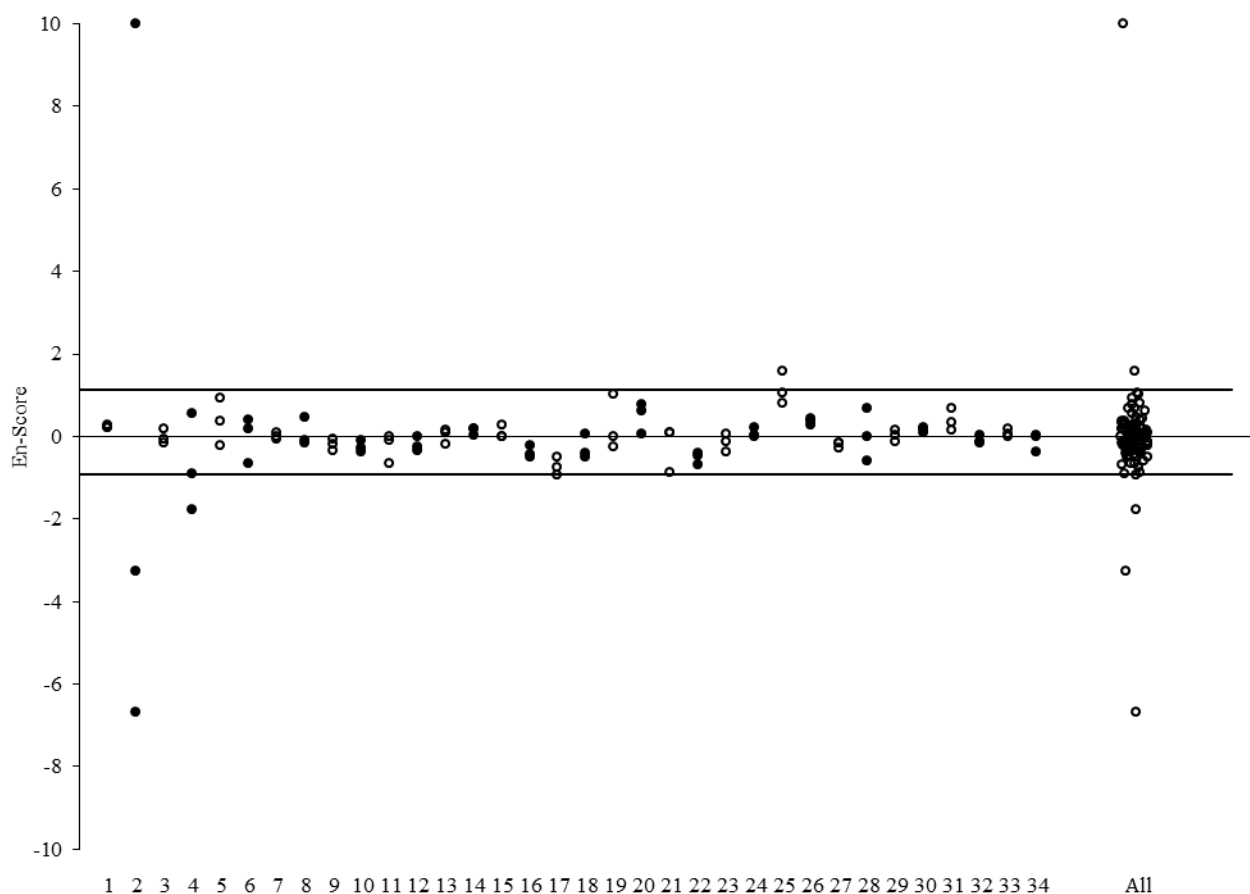


Figure 6 Summary of participants' E_n-Score

Ninety-five of 102 numeric results (93%) returned a satisfactory E_n-score with $|E_n| \leq 1$.

- Thirty-one (91%) – **1, 3, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 26, 27, 28, 29, 30, 31, 32, 33** and **34** returned satisfactory scores for all three samples;
- Two laboratories returned at least one questionable E_n-score; and
- Laboratory **2** returned $|E_n| > 1$ for all samples.

6.5 Identification of Cutting Agent

Samples were prepared using an illicit seizure of cocaine hydrochloride, approximately 84% base (m/m) supplied by the Australian Federal Police. The study coordinator added, phenacetin in Sample S2 and procaine in Sample S3.

Thirty-three participants (96%) reported on the identity of the cutting agents and correctly identified all of them. (Table 8).

6.6 Theoretical Concentration (% base cocaine)

The maximum concentration of cocaine as base (MW = 303.3) in anhydrous cocaine Hydrochloride (MW = 339.8) is 89.3%. Laboratory **25** reported the result for sample S1 as 95% base m/m.

6.7 Participants' Analytical Methods

Participants were requested to analyse the samples using their normal test methods and to report a single result for each sample as they would normally report to a client. Results reported in this way reflect the true variability of results reported to laboratory clients. The method descriptions provided by participants are presented in Table 1.

A summary of accreditation status, participants' methods and reference standards is presented below.

| Accredited | Laboratory Code |
|------------------|---|
| Yes to ISO 17025 | 1 3 4 5 6 7 8 9 10 11 13 14 15 16 17 18 19 20 22 23 24 25 26 27 28 30 31 32 34 |
| Yes to other | 7 23 34 |
| No | 2 12 21 29 33 |

| Sample Mass Used (mg) | Laboratory Code |
|-----------------------|--|
| 4-10 | 3 14 19 33 |
| 11-30 | 4 8 9 10 12 13 15 16 17 18 20 21 24 25 26 29 30 31 32 |
| 31-50 | 1 2 5 6 7 22 23 27 28 34 |
| 51-100 | |
| 101-150 | 11 |

| Instrument Used for quantification | Laboratory Code |
|------------------------------------|---|
| GC-FID | 6 9 13 16 19 20 22 23 28 30 |
| UPLC-MS(MS) | 1 2 18 25 26 27 |
| HPLC (UPLC)-DAD | 3 4 5 7 8 10 11 12 14 15 21 24 29 31 32 33 34 |
| QNMR | 17 |

| Sources of Calibration Standard | Laboratory Code |
|---------------------------------|-----------------------------------|
| NMI Australia | 2 9 16 18 22 23 25 26 27 32 |
| Lipomed | 5 7 13 20 29 31 34 |
| Sigma Aldrich | 14 15 30 33 |
| Other | 1 3 4 6 8 10 11 12 17 19 21 24 28 |

Plots of measurement extraction solvent vs z-score, measurement instrument used vs z-score and calibration standard vs z-score are presented in Figures 7, 8 and 9. No trends were observed.

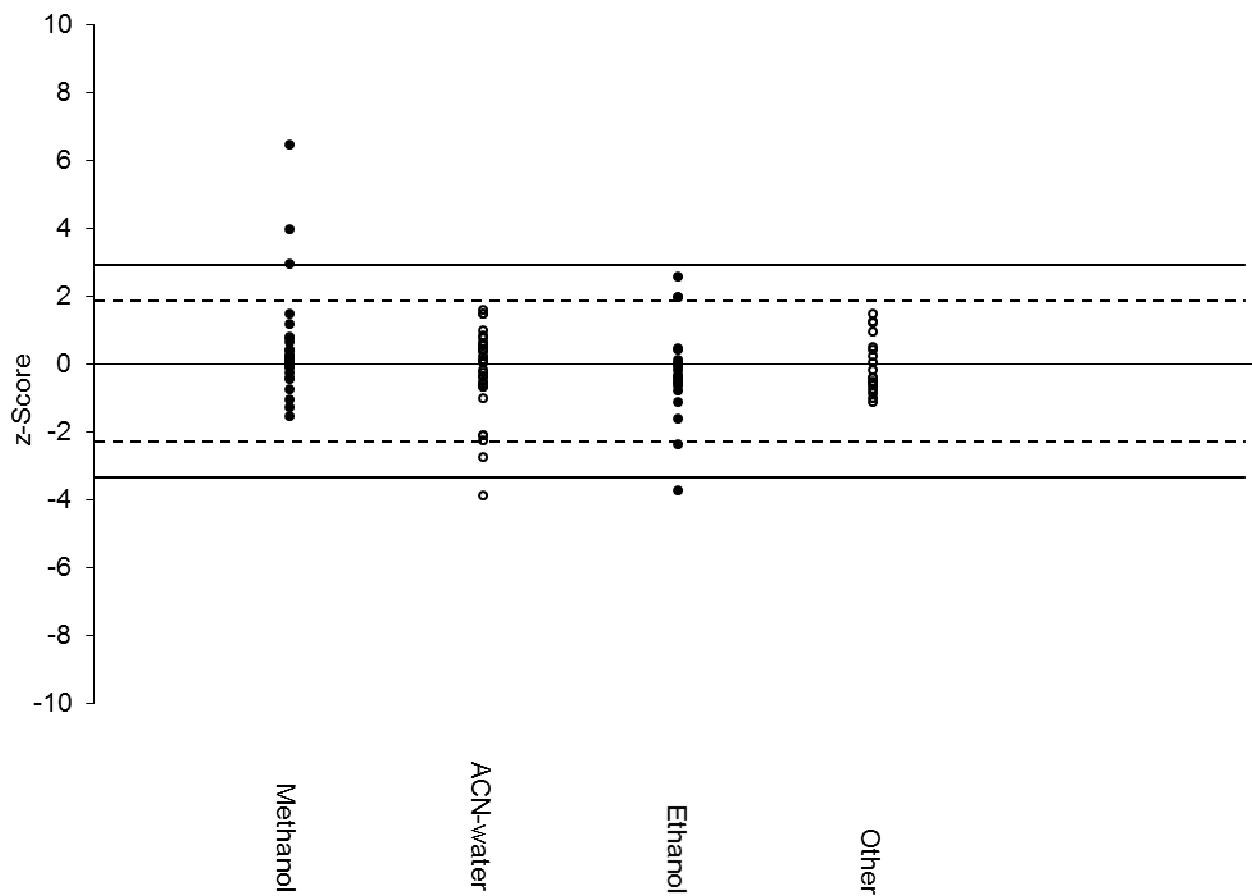


Figure 7 Extraction solvent vs z-score

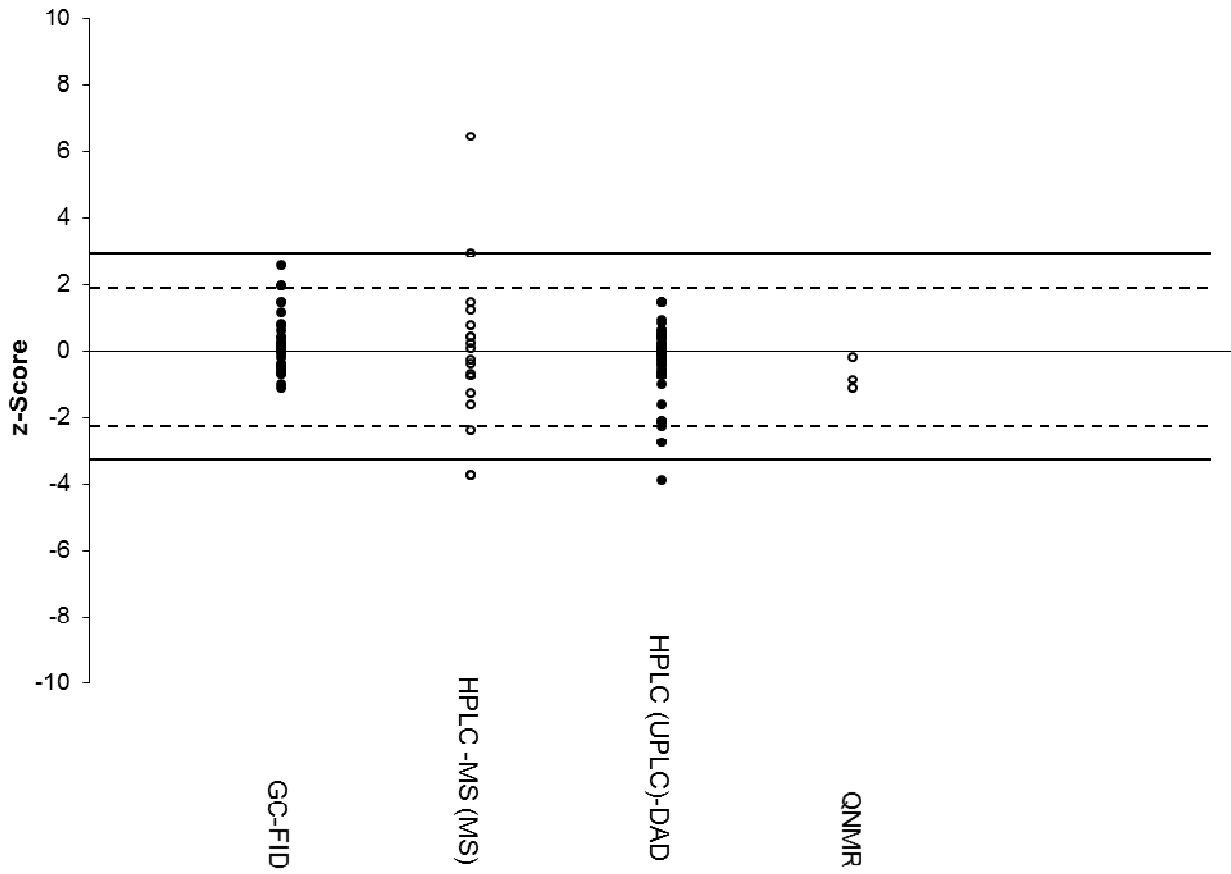


Figure 8 Measurement instrument vs z-score

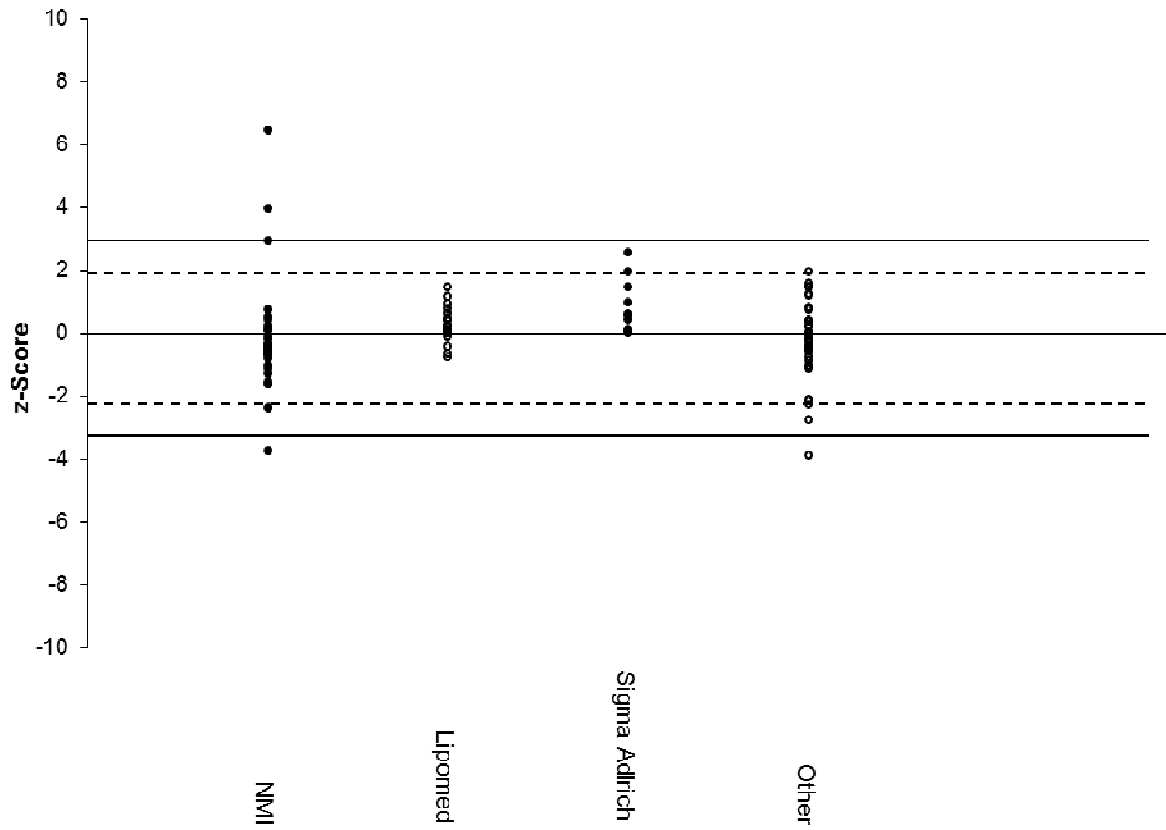


Figure 9 Calibration standard vs z-score

6.8 Summary of participation and performance in Cocaine Studies

Overall percentages of satisfactory z-scores and E_n -scores obtained by laboratories since 2009 are presented in Figure 10. The proportion of satisfactory z-scores and E_n -scores over 9 years on average is 79% and 80% respectively.

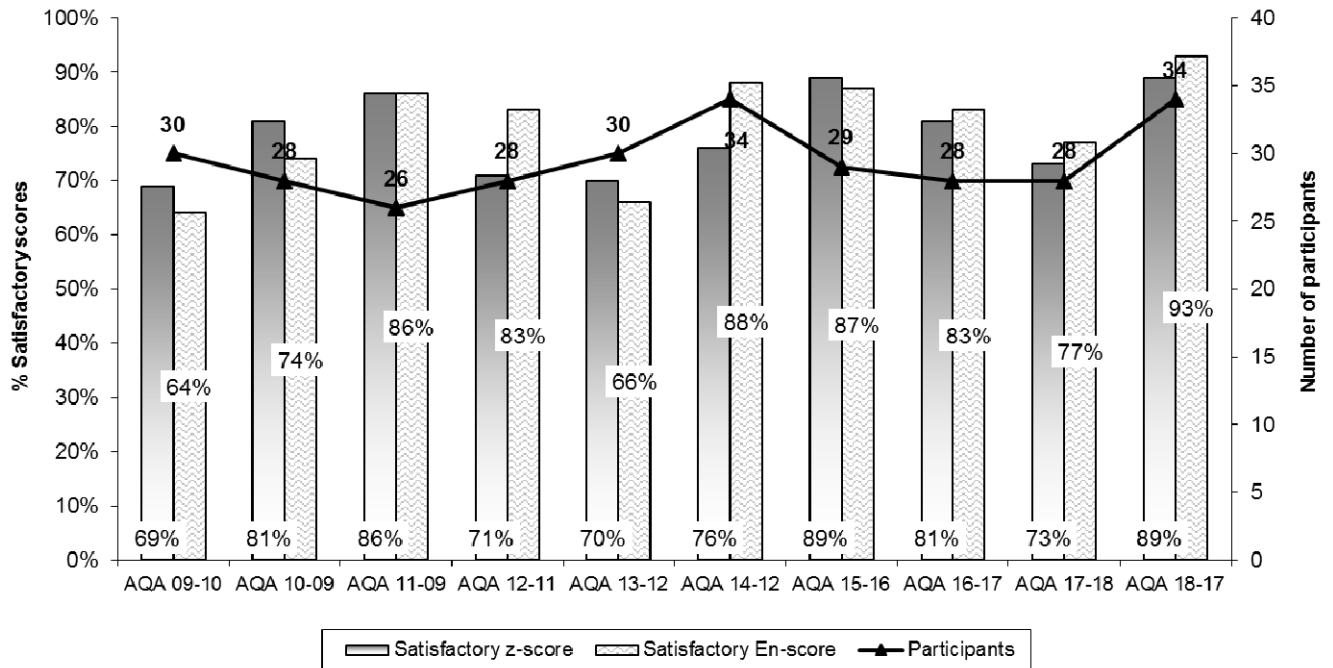


Figure 10 Summary of participants' performance since 2009

7 REFERENCES

- [1] ISO/IEC 17043:2010, Conformity assessment – General requirements for proficiency testing, ISO Geneva
- [2] NMI Chemical Proficiency Testing Study Protocol
<http://www.measurement.gov.au> → Products and Services → Chemical Proficiency Testing → Details of Our Program
- [3] NMI Chemical Proficiency Testing Statistical Manual
<http://www.measurement.gov.au> → Products and Services → Chemical Proficiency Testing → Details of Our Program
- [4] Thompson, M. E, S. L. R. and Wood, R., The international harmonized protocol for proficiency testing of (chemical) analytical laboratories, Pure Appl. Chem. 78, 145-196, 2005.
- [5] ISO/IEC 17025:2017 General requirements for the competence of testing and calibration laboratories, ISO Geneva
- [6] Eurachem/CITAC Guide Quantifying uncertainty in analytical measurement third edition, (2012), [http:// http://eurachem.org/images/stories/guides/pdf/quam2012_P1.pdf](http://eurachem.org/images/stories/guides/pdf/quam2012_P1.pdf)
- [7] ISO 13528:2015, Statistical methods for use in proficiency testing by interlaboratory comparisons, ISO Geneva
- [8] Thompson, M. and Lowthian, P.J., A Horwitz-like function describes precision in a proficiency test, Analyst, 120, 271-272, 1995.

APPENDIX 1 - MEASUREMENT UNCERTAINTY OF THE ASSIGNED VALUE

When the assigned value is calculated as the robust average using the procedure described in 'ISO13528:2015, Statistical methods for use in proficiency testing by interlaboratory comparisons – Annex C'⁷, the uncertainty is estimated as:

$$u_{\text{rob average}} = 1.25 * S_{\text{rob average}} / \sqrt{p} \quad \text{Equation 4}$$

where:

| | |
|--------------------------|-------------------------------------|
| $u_{\text{rob average}}$ | robust average standard uncertainty |
| $S_{\text{rob average}}$ | robust average standard deviation |
| p | number of results |

The expanded uncertainty ($U_{\text{rob average}}$) is the standard uncertainty multiplied by a coverage factor of 2 at approximately 95% confidence level.

A worked example is set out below in Table 10.

Table 10 Uncertainty of assigned value for Sample S1 as % base (m/m)

| | |
|--------------------------|-------|
| No. results (p) | 34 |
| Robust average | 84.89 |
| $S_{\text{rob average}}$ | 2.07 |
| $u_{\text{rob average}}$ | 0.44 |
| k | 2 |
| $U_{\text{rob average}}$ | 0.88 |

The assigned value for Sample S1 is $84.9 \pm 0.9\%$ cocaine base (m/m).

APPENDIX 2 - ACRONYMS AND ABBREVIATIONS

| | |
|------------------------|---|
| ASCLD | American Society of Crime Laboratory Directors |
| CITAC | Cooperation on International Traceability in Analytical Chemistry |
| CRM | Certified Reference Material |
| CV | Coefficient of Variation |
| DAD | Diode Array Detector |
| $ E_n $ | Absolute value of an E_n -score |
| FID | Flame Ionization Detector |
| GC | Gas Chromatography |
| GC-MS | Gas Chromatography Mass Spectrometry |
| GUM | Guide to the expression of uncertainty in measurement |
| HPLC | High Performance Liquid Chromatography |
| ISO | International Standards Organisation |
| LC | Liquid Chromatography |
| Max | Maximum value in a set of results |
| Md | Median |
| Min | Minimum value in a set of results |
| NATA | National Association of Testing Authorities |
| NMI | National Measurement Institute Australia |
| NR | Not Reported |
| NT | Not Tested |
| PDA | Photodiode array |
| PT | Proficiency Test |
| QNMR | Quantitative Nuclear Magnetic Resonance |
| Robust CV | Robust Coefficient of Variation |
| Robust SD | Robust Standard Deviation |
| SI | International System of Units |
| Target SD (σ) | Target standard deviation |
| UPLC | Ultra Performance Liquid Chromatography |
| UV | Ultraviolet |
| $ z $ | Absolute value of a z-score |

END OF REPORT